

Roelfes, G.; Feringa, B. L. Angew. Chem., Int. Ed. Engl. **2005**, 44, 3230

Michel Grenon June 25<sup>th</sup>, 2005

## **Presentation Outline**

#### **Deoxyribozymes (DNAzymes): DNA Catalysts for Bioorganic Chemistry**

Example of DNAzyme that cleaves RNA

In vitro selection approach to synthesize DNAzymes

Example of DNAzyme that ligates RNA

Other reactions catalyzed by DNAzymes

DNAzymes catalytic parameters, mechanism and structures

#### **DNA-based Asymmetric Catalysis**

Concept

Synthesis of ligands

Application to <sup>a</sup> copper-catalyzed Diels-Alder reaction

**Perspectives** 

 $\triangleright$  Relatively few studies focus on nucleic acids as catalysts for bioorganic chemistry

- The study of DNAzymes is only about a decade old, whereas that of RNAzymes goes back over 20 years
- $\triangleright$  Reasons for lack of development in this field
	- Compared with proteins, there are much less functional groups available



Emerging Area: Silverman, S. K. Org. Biomol. Chem. **2004**, 2, 2701

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**base**

 $\triangleright$  First examples of catalytic DNA: deoxyribozymes that cleave RNA



- This reaction is the same as that promoted by most protein ribonucleases such as R Nase A
- An *in vitro* selection approach can be used to identify RNA-cleaving DNAzymes

 $\triangleright$  In vitro selection approach to synthesize DNAzymes that cleave RNA



! Other examples of catalytic DNA: deoxyribozymes that ligate RNA



 $\triangleright$  Deoxyribozyme catalytic parameters, mechanism and structures

Quantitative assessment of a DNAzyme's catalytic activity can be made by comparing its rate constant to that of an appropriate background reaction

rate enhancement =  $k_{\text{obs}}/k_{\text{bkg}}$ 

rate enhancements of  $10^6$  to  $10^7$  for RNA ligation reactions rate enhancement as high as  $10^{10}$  observed for other DNAzymes

Little is known about the structures and mechanisms of any DNAzymes

 $\triangleright$  Why study DNAzymes instead of RNAzymes?

- If DNA and RNA have similar catalytic potential, practical concerns favor the use of DNA;
	- 1) DNA less expensive to make by solid-phase synthesis (ca 7 times less)
	- 2) DNA can generally be made in longer sequence lenghts and higher purity
	- 3) Relative chemical and biochemical stability (ubiquitous ribonucleases)

 $\triangleright$  Other reactions catalyzed by DNAzymes that covalently modify nucleic acids

Change in the phosphorylation status of an RNA or DNA strand

DNA phosphorylation DNA adenylation (capping)

- DNA deglycosylation
- Porphyrin metalation
- Thymine dimer photoreversion
- DNA cleavage

 $\triangleright$  What do all these processes have in common?

The use of *single-stranded* DNA for catalysis

 $\geq$  Is it possible to use duplex-DNA to catalyze a specific reaction?



 $\triangleright$  Can the chirality of the DNA double helix be transfered directly to a metal-catalyzed reaction?

Exploit the propensity of small aromatic molecules to interact with DNA in a noncovalent, yet kinetically stable way



• The reaction...



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 $\triangleright$  Synthesis of the ligands



#### $n = 3$

 $R = Me$ ,  $t$ -Bu, Benzyl, 1-Naphthylmethyl, 2-Naphthylmethyl 4-MeOC $_6$ H<sub>4</sub>CH<sub>2</sub>, 3,5(MeO)<sub>2</sub>C $_6$ H<sub>3</sub>CH<sub>2</sub>

#### $n = 2, 4, 5$

 $R = 1$ -Naphthylmethyl, 3,5(MeO)<sub>2</sub>C<sub>6</sub>H<sub>3</sub>CH<sub>2</sub>

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<sup>a</sup> Catalyst (0.18 mM), dienophile (4 mM), cyclopentadiene (34 mM)

 $<sup>b</sup>$  Calf thymus DNA  $<sup>c</sup>$  ca. 50% conversion</sup></sup>

• No significant ee when  $R = 2$ -Naphthylmethyl

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<sup>a</sup> Catalyst (0.18 mM), dienophile (4 mM), cyclopentadiene (34 mM)

b Calf thymus DNA

 $\text{c}$  DNA = synthetic duplex d(GACT)<sub>2</sub>-(AGTC)<sub>2</sub> (0.39 mM), cyclopentadiene (21 mM)

 $d$  ca. 50% conversion

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<sup>a</sup> Reaction performed at 5 ˚C



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# **Perspectives**

- $\triangleright$  The chirality of duplex DNA can be transfered directly to a catalytic reaction
- $\triangleright$  *Both* enantiomers of the Diels-Alder adduct are accessible by a judicious choice of ligand
- $\triangleright$  Rapid structural variation and optimization of catalysts for new reactions
- ! Ease of purification (Cu-ligand-DNA complex remains in aqueous solution)

Futur work should focus on

- $\triangleright$  The possibilty to address specific DNA sequences by using a selective DNA binding moiety tethered to the catalyst
- $\triangleright$  Extending to other reactions that can be performed in buffered aqueous solutions (metal-catalyzed reactions, organocatalysis)